

Available online at http://bjas.bajas.edu.iq https://doi.org/10.37077/25200860.2024.37.2.18 College of Agriculture, University of Basrah

Basrah Journal of Agricultural Sciences

ISSN 1814 - 5868

Basrah J. Agric. Sci., 37(2), 240-248, 2024 E-ISSN: 2520-0860

### Genetic Relationships Analysis of Some various Species Using Cytochrome b Genes as a Phylogenetic Marker Lajan Salahaldin Ahmed

Department of Animal Resources, College of Agriculture Engineering Science, Salahaddin University-Erbil \*Corresponding author email: Lajan.ahmed@su.edu.krd

Received 21<sup>st</sup> September 2024; Accepted 27<sup>th</sup> November 2024; Available online 31<sup>st</sup> December 2024

**Abstract:** This study examined the genetic relationships among six species—chicken (*Gallus gallus*), quail (Coturnix coturnix), duck (Anas platyrhynchos), goose (Anseranser domestica), and turkey (Meleagris gallopavo) and rabbit (Lepus capensis), using partial mitochondrial cytochrome b (Cyt-b) gene sequences. PCR amplification of the mitochondrial Cvt-b gene resulted in an approximately 358 bp amplicon in size for all species, and sequences alignment and phylogenetic analyses were performed with Bio Edit and MEGA X software's to determine genetic similarities and distances. Results revealed high nucleotide similarity between chicken and quail (89.1%) and chicken and turkey (83.6%). The closest genetic relationship was between duck and domestic goose, with 87.4% identity and the smallest genetic distance of 0.126. Chicken (Gallus gallus) demonstrated a relatively close relationship with both duck and domestic goose, with identities (83.6%). While, Turkey (Meleagris gallopavo) exhibited a slightly more distant relationship with avian species, with the lowest similarity at 81.6% with domestic goose. The rabbit, positioned at the root of the phylogenetic tree, exhibited the greatest genetic distance from avian species, with a 28.3% distance and 71.7% identity with chicken. This study provides solid evidence of the effectiveness of using Cyt-b gene sequences as a reliable tool for species identification across various applications, underscoring the effectiveness of partial Cyt-b gene sequences for avian species identification and genetic analysis, which is valuable for breeding and diversity studies.

Keywords: Cyt-b gene, genetic distance, phylogenetic tree, sequence analysis, various species

### Introduction

Avian species, such as chickens, quails, turkeys, ducks, and geese, are valuable domesticated birds raised by humans for their eggs and meat (Farrell, 2014). They provide high-quality animal proteins and are often included in various meat products. However, substitution of these meats with lower-cost alternatives has led to fraudulent practices to increase financial gains. Such adulteration undermines fair competition and disregards consumer interests (Abbas *et al.*, 2019; Bohme *et al.*, 2019).To enforce legislation aimed at protecting endangered bird species, it is crucial to accurately identify avian species. DNA testing is often employed for this purpose, particularly when morphological information is limited, such as in cases involving only feather fragments

powdered remains. Sequence-based or identification of cytochrome b genes in mitochondria is the most popular exploited among DNA-based techniques (Hebert et al., 2003; Lee et al., 2008). Accurately identifying a species is also an important element of the studies of biodiversity (Ardura et al., 2011). Other approaches that have been applied for species identification include but are not limited to morphological systems, features, immune and their techniques, electrophoresis, gel and chromatography (Taylor *et al.*, 1993: Andrasko & Rosen, 1994; Espinoza et al., 1999; Czesny et al., 2000). Many recent identification approaches have particularly failed to separate some closely related bird species and therefore, approaches with better resolution and sensitivity are needed (Bellis et al., 2003). Molecular genetic analysis, particularly using mitochondrial DNA (mt-DNA) sequences, offers a promising solution. Mitochondrial DNA, due to its simpler structure compared to genomic DNA, maternal inheritance, and lack of recombination in vertebrates, makes its sequence more conserved (Rokas et al., 2003). The rate of base substitution in mt-DNA is approximately 5 - 10 times higher than in the nuclear genes which enables the differentiation of a wide range of bird species even those that are closely related in the same families and genera. This is so as a result of over some time the number of base substitutions increases (Russell et al., 2000). These scenarios point out that lineage with differential mt-DNA is a useful source for species identification and characterization in both taxonomy and phylogenetic research

(Noro et al., 1998; Olschewsky & Hinrichs, 2021). Cytochrome b (Cyt-b) proteins are favored by biologists for their active taxonomic involvement in the and phylogenetic analysis of organisms at various levels, including species and family, etc. The mt-cyb gene has been functional in testing distances among taxa, short of the order among several groups of animals. The mt-Cvb gene offers information specific to the species concerned within the purported specimens whose forensic applications range from the development of the phylogenetic tree to forensic caseworks investigations including source determination of the biological matter within the caseworks (Yacoub et al., 2015; Hartatik et al., 2019). The goal of this study was to use cytochrome b (Cvt-b) sequenceing as a reliable tool for species identification.

# **Materials and Methods**

## **Avian Species Blood Samples**

Ten blood samples from each of the following species: chicken (Gallus gallus), quail (Coturnix coturnix), duck (Anas platyrhynchos), goose (Anseranserdo mestica), turkey (Meleagris gallopavo), and rabbit (Lepus capensis), were collected for molecular analysis. Blood was drawn from the jugular vein using a 10 ml syringe. Prior to collection, the area was cleaned, hair was trimmed, and the site was sterilized with 70% ethanol. The samples were positioned in test tubes that included EDTA and were held at -20°C until the DNA extraction procedure was performed.

### **DNA Isolation**

The DNA was extracted from blood samples of eight bird species using a GeNet Bio, Korea blood DNA extraction kit. Following the manufacturer's instructions, the extracted DNA's quality and quantity were determined using a Nanodrop 1000 spectrophotometer (UK) and a 1% agarose gel electrophoresis method, respectively.

### PCR and gel electrophoresis

methodology of experiment The the involved two primers that were specifically chosen to target and amplify a segment of the mitochondrial cytochrome b (Cyt-b) gene:(5'CCATCCAACATCTCAGCATGAT GAAA3')and(5'CCCCTCAGAATGATATT TGTCCTCA-3'), as indicated by (Awad et al., 2015). The PCR reaction was carried out in a total volume of 20 µL, which included a mixture of 10 µL of Green Master Mix (featuring 25 units/mL of Taq DNA polymerase, 200 µM of each dNTP, and 1.5 mM of MgCl<sub>2</sub>), 2 µL of genomic DNA, 1 µL of each primer (at a concentration of 10  $\mu$ M), and 6  $\mu$ L of DNase-free water.The PCR amplification was conducted utilizing a Tprofessional thermo cycler (Biometra, Germany), under the specified thermal conditions: an initial denaturation step at 95°C for 10 minutes, followed by 33 cycles consisting of denaturation at 95°C for 30 seconds, annealing at 50°C for 40 seconds, and extension at 72°C for 50 seconds. The amplification concluded with a final extension at 72°C for 6 minutes. Afterward,  $10 \ \mu L$  of the PCR products were applied to a

2% agarose gel, which was stained with Safe Dye (Catalog No. B-2010, GeNet Bio, Korea) and viewed under ultraviolet (UV) light. The gel images were captured utilizing a Gel Documentation System (Bio Doc. Analyse, Biometra, Germany).

## **DNA** sequencing

The PCR products were sequenced using the set of amplifying primers, both forward and reverse. This particular sequencing was performed by Macrogeninc, Inc. a South Korean commercial company offering both sequencing and purification services.

# Aligning Sequences and Evolutionary Tree Construction

The amplified mitochondrial cytochrome b (Cyt-b) gene sequences were aligned with http://www.ncbi. BLAST available at nlm.nih.gov/BLAST, and then retrieved mitochondrial Cvt-b genes of closely related species from GenBank. The genetic diversity among accessions was estimated by using the MEGA X software (version 10.2.6) (Kumar et al., 2018) with multiple sequence alignment file. Counting the number of base pair substitutions between sequences and using Kimura's 2-parameter model to correct for evolutionary rates, we calculated genetic distances (d) for each locus among the accessions. Phylogenetic tree construction was based on the maximum likelihood (ML) method with the neighbor-joining algorithm. Figure 3 shows that 1,000 bootstrap replicates were conducted to test the reliability of the resulting phylogenetic trees (Tripathi et al., 2013).

	understudy and also retrieved from Gen Bank.						
	Taxonomic name	Popular name	Accession No				
1-	Coturnix coturnix	quail	PQ356919				
2-	Gallus gallus	chicken	PQ356920				
3-	Anseranser domestica	duck	PQ356917				
4-	Anas Platyrhynchos	goose	PQ356918				
5-	Meleagris gallopavo	turkey	PQ356922				
6-	Lepus capensis	rabbit	PQ356921				

 Table (1): Phylogenetic tree, based on the mitochondrial cytochrome b gene of avian species understudy and also retrieved from Gen Bank.

### **Results and Discussion**

The investigation revealed that using universal primers for PCR amplification targeting a segment of the mitochondrial cytochrome b (Cytb) gene resulted in a single PCR product. This product was analyzed using a 2% agarose gel, revealing a fragment size of approximately 358 base pairs (bp) across the six various species examined, as shown in (Fig. 1).

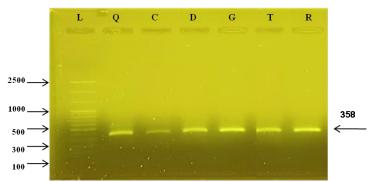


Fig.(1): The *mt-Cytb* universal primer was employed to amplify PCR products from various species.Lane L: 100 bp DNA marker, Lane Q is quail, Lane C is chicken, Lane D is duck, Lane G is goose, Lane T is turkey, and Lane R is rabbit.

The study involved submitting mitochondrial cytochrome b (Cyt-b) gene sequences from various species chickens (Gallus- gallus), quails (Coturnix- coturnix), ducks (Anasplatyrhynchos). (Anseransergeese domestica), turkeys (Meleagris- gallopavo), and rabbits (Lepus-capensis)to Gen Bank under accession numbersPQ356920, PQ356918, PO356919. PQ356917, PQ356922 and PQ356921. The Bio Edit software (version7.2.5) was utilized to align

and compare these sequences with those of other species. The partial sequencing analysis of the *mt-Cytb* gene, excluding primer regions, was conducted over a length of 280 base pairs (*bp*), as detailed in (Table 1 and Fig. 2). These results are consistent with the observations reported by Abdul-Hassan & Tauma, (2014) as well as (Awad *et al.*, 2014).

In a comparative analysis of the *mt-Cytb* gene sequences from five bird species

alongside *Lepus capensis*, it was determined that the nucleotide similarity was 89.1% between chicken (*Gallus gallus*) and quail (*Coturnix coturnix*), and 83.6% between chicken (*Gallus gallus*) and turkey (*Meleagris gallopavo*). In contrast, the nucleotide identity of rabbit (*Lepus capensis*) was found to be lower when compared to the avian species under

investigation. Furthermore, Sequencing generally involves examining part or all of mitochondrial genome the and then comparing it to existing sequences in Gen Bank (NCBI). The nucleotide similarity sequenced mitochondrial between the genomes and the reference sequences for the same species varied between 98.22% and 99.66%.

		10	20	30	40	50	60	70
Gallus gallus	TCATGAC	CCGAATCCTC	AACCGGCCTA	CTACTAGCCA	TGCACTACAC	AGCAGACACA	TCCCTAGCCI	TCTC
Coturnix coturnix				•••••				
Anas Platyrhynchos	.GG.C	ATT	C	G	.A	c	AC	• • • •
Anser anser domestica	.AGCC	A.AA	AG	•••••	.A	ст		• • • •
Meleagris gallopavo	c!	Т.АТ.А	c	•••••	.AT	<b>T</b> C	A.TTA.	• • • •
Lepus capensis	.GC.T	T.ATT.A	CT	T.CT	.A	CT.T	A.AACA.	••••
	70	80	90	100	110	120	130	140
Gallus gallus	FCCTCCGT	AGCCCACACT	TGCCGGAAC	STACAATACGG	CTGACTCAT	CGGAATCTCC	ACGCAAACG	GCGCC
Coturnix coturnix		A	<b>T</b> A	G		2c	.T	A
Anas Platyrhynchos	A	GTAA	<b>T</b> A				т.	
Anser anser domestica	A	<b>T</b> A	AG		A	cc	c	.т. т
Meleagris gallopavo	тт	GTA	A		TC.	TC	.TGT.	
Lepus capensis	AG	.AT.T.	AG.T.	TA.CT	T	AT.C	т	.AA
	140	150	160	170	180	190	200	210
Gallus gallus	CCTCATTC	TTCTTCATCT	GTATCTTCCI	TTCACATCGGA	CGAGGCCTA	TACTACGGCT	CTACCTCTA	CAAGO
	.A		.c	.c			T	A.
				ATT				
Anser anser domestica								
Meleagris gallopavo			.c	AT	c	.TTT.	GA!	тА.
Lepus capensis	A. TA.A	TT.	.CCA.A.	.AG.AC	CCAA.C	TA	AACA	.CTA.
	210	220	230	240	250	260	270	280
Gallus gallus	GGAAACC	TGAAACACAG	GAGTAATCCT	CCTCCTCACA	CTCATAGCCA	CCGCCTTTGT	GGGCTATGTT	CTCCC
Coturnix coturnix	A			GT		.TTC	AACC	т.А.
Anas Platyrhynchos	A	т	G	AG.C	GA.	.AC	AC	G
	_							7
Anser anser domestica	A	• • • • • • • • • • •		AC	AA.	.TC	a	· · · · · ·
Anser anser domestica Meleagris gallopavo				AC				

# Fig. (2): The Bio Edit software was used to align the partial mitochondrial cytochrome b (*mt-Cytb*) gene sequences from chicken (*G. gallus*), quail (*C. coturnix*), duck (*A. platyrhynchos*), goose (*A. anserdomestica*), turkey (*M. gallopavo*), and rabbit (*L. capensis*), with sequence identities represented by dots.

The results indicate that *Anas platyrhynchos* (Duck) and *Anseranser domestica* (Domestic

Goose) share the closest genetic relationship, with a high percentage of identity (87.4%)

and the smallest genetic distance (0.126). Gallus gallus (Chicken) sequence also shows a relatively close genetic relationship with both Anas platyrhynchos (Duck) and Anseranser domestica (Domestic Goose), with a percentage of identity around 83.6%. The derived sequence Meleagris gallopavo (Turkey) has a slightly more distant relationship with the other avian species, with percentage identities ranging from 84.6% with those of Anas platvrhynchos (Duck) and Coturnix coturnix, but showed less similarity 81.6% with Anseranser domestica (Domestic Goose). The genetic distances among the avian species are generally low, reflecting their closer evolutionary relationships. In contrast, Lepus capensis shows significantly greater genetic

distances and lower percentage identities with the avian species, emphasizing its distinct evolutionary lineage.

The cytochrome b (*Cyt-b*) gene has been fully or partially sequenced in a wide range of animals, including birds, mammals, fish, amphibians, reptiles (Chow *et al.*, 1993; Ram *et al.*, 1996; Quinterio *et al.*, 1998; Lindstrom, 1999; Parson *et al.*, 2000), as well as some invertebrates (Lee *et al.*, 2009). Additionally, *Cyt-b* gene sequences have been utilized to identify meats and meat products from birds like song thrush, quail, and sparrow, and other species such as red deer, roe deer, Pyrenean ibex, and chamois (Chikuni *et al.*, 1994; Matsunaga *et al.*, 1998; La Neve *et al.*, 2008).

 Table (2): The genetic distance is represented below the diagonal, while the percentage of identity among the various species analyzed is illustrated above the diagonal.

No.	Species	1	2	3	4	5	6
1	Gallus gallus		89.1	82.9	83.6	83.6	71.7
2	Coturnix coturnix	0.109		84.3	86.0	84.6	70.6
3	Anas Platyrhynchos	0.171	0.157		87.4	84.6	71.0
4	Anseranser domestica	0.164	0.140	0.126		81.6	73.4
5	Meleagris gallopavo	0.164	0.154	0.154	0.184		71.0
6	Lepus capensis	0.283	0.294	0.290	0.266	0.290	

The greatest genetic distance observed is between *Lepus capensis* and *Gallus gallus* (Chicken) at 28.3%, with the percentage of identity being the lowest at 71.7%. The table 2 highlights the closer genetic relationships among the avian species compared to the more distantly related *Lepus capensis*. The genetic distance ranged from 0.109 to 0.294 among various species as shown in Table 2 below the diagonal. Awad *et al.*, (2014) conducted a comparative analysis of *mt-Cytb* gene sequences from five bird species, revealing nucleotide similarity percentages of 88.60% between the chicken (*Gallus-gallus*) and the Japanese quail (*Coturnix-japonica*), as well as 86.64% between the laughing dove (*Spilopelia-senegalensis*) and the rock pigeon (*Columba livia*). Conversely, the sequences of the Muscovy duck (*Cairina-moschata*) demonstrated a lower degree of sequence identity than the

other avian species included in the study. Dave *et al.*, (2021) conducted a comparison between two indigenous chicken populations, Aravali and Ankleshwar, with various domestic chicken breeds. They concluded that the Red Jungle Fowl and Gray Jungle Fowl are the wild species that all domestic chicken breeds descend from.

The phylogenetic tree evaluates the relationships among several bird species (*Gallus gallus, Coturnix coturnix, Meleagris gallopavo, Anas platyrhynchos,* and *Anseranser domestica*) and one mammal

(Lepus capensis) using genetic data. The tree reveals that Gallus gallus (domestic chicken) and Coturnix coturnix (common quail) share the most recent common ancestor, with Meleagris gallopavo (wild turkey). These three species form a closely related group. Anas platyrhynchos (mallard duck) and Anseranser domestica (domestic goose) also form a close relationship but they are less closely related to the first three species. Although Lepus capensis is a mammal, it appears at the base of the phylogenetic tree due to its substantial genetic distance from the avian species.

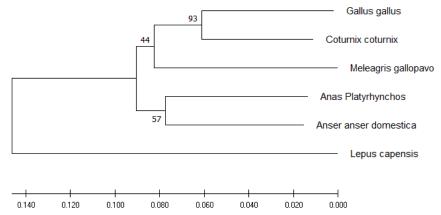


Fig. (3): Analysis of cytochrome b gene sequences was performed on species collected for the construction of a phylogenetic tree using maximum likelihood (ML) analysis.

Genetic distances among various poultry breeds or genotypes can provide an accurate picture of genetic diversity (Ahmed, 2020; Ahmed & Al-Barzinji, 2019). This type of information is important for breeders, particularly in designing mating systems for crossbreeding

## Conclusion

The study investigated species differentiation by amplifying and sequencing a specific fragment of the mitochondrial cytochrome b (mt-Cytb) gene. The mitochondrial Cyt-b gene, known for its high

genetic variability across species, was chosen as it provides a reliable molecular for species identification in phylogenetic studies.

# ORCID

L.S.A., https://orcid.org/0000-0002-3696-1108

# **Conflict of interest**

The authors affirm that this work has been submitted without any conflicts of interest.

# References

- Abbas, A., Abbas, R.Z., Khan, M.K., Raza, M.A., Mahmood, M.S., Saleemi, M.K., Hussain, T., Khan, J.A. & Sindhu, Z.U.D. (2019).
  Anticoccidial effects of Trachyspermumammi (Ajwain) in broiler chickens. *Pakistan Veterinary Journal*, 39(2), 301-304. https://doi.org/10.29261/pakvetj/2019.056
- Abdul-Hanssan, I.A., & Tauma, J.A. (2014).
  Identification of some meat species using PCR and Multiplex PCR of Mitochondrial Cytochrome B Gene. *Iraqi poultry sciences journal*, 8(1),1-9.
- Ahmed, L.S. (2020). DNA Markers of Three Genes and Their Associations with the Meat and Egg Production Traits in Local Varieties of Quail in Kurdistan Region. Doctoral dissertation, Department of Animal Production, College of Agriculture Engineering Sciences, Salahaddin University-Erbil.Iraq.
- Ahmed, L.S., & Al-Barzinji, Y.M.S. (2019). Genetic Diversity among Local Quail Using RAPD-DNA Marker. *Revista Cientifica de la Facultade de Veterinaria*, 29(3).
- Andrasko, J. & Rosen, B. (1994). Sensitive identification of hemoglobin in bloodstains from different species by high performance liquid chromatography with combined UV and fluorescence detection. *Journal of Forensic Sciences*, *39*(4), 1018-1025. https://doi.org/10.1520/jfs13680j
- Andrzej D., & Knapik, K. (2005). A new PCR-RFLP within the domestic pigeon (Columba livia var. domestica) cytochrome b (MTCYB) gene. Journal of Applied Genetics, 46(3),315-317.
- Ardura, A., Planes, S., & Garcia-Vazquez, E. (2011). Beyond biodiversity: fish metagenomes. *PLoS One*, 6(8), e22592. https://doi.org/10.1371/journal.pone.0022592
- Awad, A., Khalil, S.R., &Abd-Elhakim, Y.M. (2014). Molecular phylogeny of some avian species using Cytochrome b gene sequence analysis. *Iranian journal of veterinary research*, 16(2), 218-222.
- Bellis, C., Ashton, K.J., Freney, L., Blair, B., & Griffiths, L.R. (2003). A molecular genetics approach for forensic animal species identification. Forensic science international, *134*(2-3), 99-108. https://doi.org/10.1016/s0379-0738(03)00128-2
- Böhme, K., Calo-Mata, P., Barros-Velázquez, J., & Ortea, I. (2019). Review of recent DNA-based methods for main food-authentication topics. *Journal of agricultural and food chemistry*, 67(14), 3854-3864. https://doi.org/10.1021/acs.jafc.8b07016
- Bravi, C.M., Lirón, J.P., Mirol, P.M., Ripoli, M.V., Peral-García, P., & Giovambattista, G. (2004).

A simple method for domestic animal identification in Argentina using PCR-RFLP analysis of cytochrome b gene. Legal Medicine, 6(4), 246-251.

https://doi.org/10.1016/j.legalmed.2004.06.003

- Chikuni, K., Tabata, T., Kosugiyama, M., Monma, M., & Saito, M. (1994). Polymerase chain reaction assay for detection of sheep and goat meats. *Meat Science*, 37(3), 337-345. https://doi.org/10.1016/0309-1740(94)90051-5
- Chow, S. (1993). PCR-RFLP analysis on thirteen western Atlantic snappers (subfamily Lutjaninae): a simple method for species and stock identification. *Fish Bull*, 91,619-627.
- Czesny, S., Dabrowski, K., Christensen, J.E., Van Eenennaam, J., & Doroshov, S. (2000). Discrimination of wild and domestic origin of sturgeon ova based on lipids and fatty acid analysis. *Aquaculture*, 189(1-2), 145-153. https://doi.org/10.1016/S0044-8486(00)00364-1
- Dave, A.R., Chaudhary, D.F., Mankad, P.M., Koringa, P.G., & Rank, D.N. (2021). Genetic diversity among two native Indian chicken populations using cytochrome c oxidase subunit I and cytochrome b DNA barcodes. *Veterinary World*, 14(5), 1389. https://doi.org/10.14202/vetworld.2021.1389-

1397

Espinoza, E.O., Lindley, N.C., Gordon, K.M., Ekhoff, J.A. & Kirms, M.A. (1999). Electrospray ionization mass spectrometric analysis of blood for differentiation of species. *Analytical Biochemistry*, 268(2), 252-261.

https://doi.org/10.1006/abio.1998.3048

- Farrell, D.J. (2014). Small-scale duck production: the way ahead. *Journal of Animal Husbandry Science and Technology*, *30*(8), 73-80.
- Hartatik, T., Hariyono, D.N.H., &Adinata, Y. (2019). Genetic diversity and phylogenetic analysis of two Indonesian local cattle breeds based on cytochrome b gene sequences. *Biodiversitas Journal of Biological Diversity*, 20(1), 17-22.

https://doi.org/10.13057/biodiv/d200103

Hebert, P.D., Cywinska, A., Ball, S.L., & DeWaard, J.R. (2003). Biological identifications through DNA barcodes. *Proceedings of the Royal Society of London. Series B: Biological Sciences*, 270(1512), 313-321.

https://doi.org/10.1098/rspb.2002.2218

Kocher, T.D., Thomas, W.K., Meyer, A., Edwards, S.V., Pääbo, S., Villablanca, F.X., & Wilson, A.C. (1989). Dynamics of mitochondrial DNA evolution in animals: amplification and sequencing with conserved primers. *Proceedings of the National Academy of Sciences*, *86*(16), 6196-6200. https://doi.org/10.1073/pnas.86.16.6196

- Kumar, S., Stecher, G., Li, M., Knyaz, C., & Tamura, K. (2018). MEGA X: molecular evolutionary genetics analysis across computing platforms. *Molecular* biology and evolution, 35(6), 1547-1549.
- La Neve, F., Civera, T., Mucci, N., &Bottero, M.T. (2008). Authentication of meat from game and domestic species by SNaPshotminisequencing analysis. Meat Science, *80*(2), 216-224. https://doi.org/10.1016/j.meatsci.2007.11.027
- Lee, J.C.I., Tsai, L.C., Huang, M.T., Jhuang, J.A., Yao, C.T., Chin, S.C., Wang, L.C., Linacre, A., https://doi.org/10.1021/jf950822t
- Rokas, A., Ladoukakis, E., &Zouros, E. (2003). Animal mitochondrial DNA recombination revisited. *Trends in Ecology & Evolution*, 18(8), 411417.
  - https://doi.org/10.1016/S0169-5347(03)00125-3
- Russell, V.J., Hold, G.L., Pryde, S.E., Rehbein, H., Quinteiro, J., Rey-Mendez, M., Sotelo, C.G., Pérez-Martin, R.I., Santos, A.T., & Rosa, C. (2000). Use of restriction fragment length

polymorphism to distinguish between salmon species. *Journal of agricultural and food chemistry*, 48(6), 21842188. https://doi.org/10.1021/jf991213e

- Taylor, A.J., Linforth, R., Weir, O., Hutton, T., & Green, B. (1993). Potential of electrospray mass spectrometry for meat pigment identification. *Meat science*, *33*(1), 75-83. https://doi.org/10.1016/0309-1740(93)90095-Y
- Tripathi, A.M., Tyagi, A., Kumar, A., Singh, A., Singh, S., Chaudhary, L.B., & Roy, S. (2013). The internal transcribed spacer (ITS) region and trnhH-psbA are suitable candidate loci for DNA barcoding of tropical tree species of India. *PloS* one, 8(2), e57934.

https://doi.org/10.1371/journal.pone.0057934

Yacoub, H.A., Fathi, M.M., &Sadek, M.A. (2015). Using cytochrome b gene of mtDNA as a DNA barcoding marker in chicken strains. *Mitochondrial DNA*, 26(2), 217223. https://doi.org/10.3109/19401736.2013.825771

### تحليل العلاقات الوراثية لبعض أنواع المختلفة باستخدام جين Cytochrome b كمؤشر النشوء

### والتطور

#### لاجان صلاح الدين احمد

قسم الثروة الحيوانية /كلية علوم الهندسة الزراعية / جامعة صلاح الدين- أربيل / العراق

المستخلص: تناولت هذه الدراسة العلاقات الوراثية بين خمسة انواع مختلفة من الطيور: الدجاح (Gallus-gallus)، والديك والسمان (Coturnix coturnix)، والبط (Anas-platyrhynchos)، والإوز (Anser-anserdomestica)، والديك الرومي (Meleagris gallopavo) فضلا عن الأرنب كمجموعة خارجية (Lepus-capensis)، وذلك باستخدام تسلسلات جزئية من جين الميتوكوندريا السيتوكروم ب (Cyt-b) نتج عن تضخيم جين الميتوكوندريا Cyt-b باستخدام تقنية

PCRحزمة بحجم 358 زوج قاعدي أساس لجميع الانواع. وتم إجراء محاذاة التسلسلات وتحليلات النشوء والتطور بواسطور المسلمة Bio Edit لتحديد أوجه التشابه والمسافات الجينية. أظهرت النتائج وجود تشابه كبير في النيوكليوتيدات بين الدجاج والسمان (89.1%) والدجاج والديك الرومي (83.6%). أقرب علاقة وراثية كانت بين البط والأوز المستأنس حيث بلغت نسبة التشابه 87.4% وأقل مسافة جينية كربية كانت 0.126. أظهر الدجاج (Gallus-gallus) علاقة جينية قريبة نسبيًا مع كل من البط والأوز المستأنس، حيث بلغت نسبة التشابه 87.4% وأقل مسافة جينية كانت مع 10.2%. في حين أظهر الدجاج كل من البط والأوز المستأنس، حيث بلغت نسبيًا مع

(gallopavo علاقة جينية أبعد قليلاً مع الأنواع الطيور الأخرى، حيث كانت أقل نسبة تشابه هي 81.6% مع الأوز المستأنس. الأرنب، الذي تم وضعه في جذر الشجرة التطورية، أظهر أكبر مسافة جينية عن الأنواع الطيور، حيث كانت المسافة 32.3% ونسبة التشابه 71.7% مع الدجاج. تسلط هذه الدراسة الضوء على العلاقات الجينية الأقرب بين الأنواع الطيور مقارنة بالأرنب، مما يبرز فعالية استخدام تسلسلات جين *Cyt*-b الجزئية في تحديد الأنواع الطيور وتحليلها جينيًا، وهو دراسة ذو قيمة في دراسات التنوع وبرامج التربية.

كلمات المفتاحية: Cyt-b جين, شجرة النشوء, تسلسلات الجينية, البعد الوراثي ,الانواع المختلفة